

# Biochemical changes in the midgut during metamorphosis in *Apis* cerana indica

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## **Abstract**

The digestive cells of midgut are responsible for the secretion of various enzymes and absorption of nutrients. During the process of metamorphosis midgut passes through, histolysis and histogenesis. As a result, destruction of larval tissue and construction of adult tissue occurs. The present work thus carried out is to know the changes that occurr in the biomolecules like DNA, RNA, Proteins, Carbohydrates etc., in relation with the remodeling of gut. Along with these biomolecules, various enzymes like amylase, invertase, protease and lipase are also estimated to know their status during metamorphosis of midgut in *Apis cerana indica*.

Keywords- Apis cerana indica, Midgut, Metamorphosis, Biomolecules.

## Introduction

Apis cerana indica is one of the indigenous species of honeybees, used for apiculture in India. The alimentary canal of the bee can be divided into three distinct regions foregut, midgut and hindgut (Snodgrass, 1935 and 1956). The midgut is the main site for the digestive processes in most of the insects (Wigglesworth, 1972 and House, 1974), the midgut epithelium is composed of columnar cells which secretes the enzymes, carries out digestion and absorption (Wigglesworth, 1977). The ultrastructural and cytochemical aspect indicates that the midgut is remodeled during metamorphosis and shows the programmed cell death in the honey bee larvae and pupae (Gregorc and Bowen, 1996 and 1997, Cruzlandim and Cavalcante 2002 and 2003, Neves et al., 2003 and Barsagade and Kelwadkar, 2008). The peptides of insect brain and midgut are suggested to have important regulatory role in digestive processes like, enzyme secretion, epithelial tissue regeneration, absorption of nutrients, working of gut musculature and maintenance of gut pH (Prabhu and Sreekumar, 1994 and Sunitha et al., 1999). In Apis cerana indica remodeling of gut occurs through histolysis and histogenesis metamorphosis but no information is available regarding biomolecule concentration and protein profile in the midgut cells during metamorphosis. Therefore, present work has been undertaken to know the changes in biomolecules, enzymes and protein profile in the midgut epithelial cells of larvae and pupae of Apis cerana indica.

# Materials and Methods Biochemical Techniques Extract Preparation:

The midguts were dissected out from the

larval and pupal stages, in ice-cold saline/Ringer's solution. The dissected midguts were homogenized using pestle and mortar for 5 minutes at room temperature in the Ringer's solution. 25 mg tissue/ml volumes of Ringer solution was added, sample was homogenized at 3000 rpm and the supernatant after centrifugation was used for estimation of total concentration of protein according to the Lowery et al., (1951) method. The total carbohydrate was estimated by Phenol-Sulphuric acid method of Dubois et al., (1962), DNA and RNA by Bruton's Diphenylamine and Dische-Orcinol methods of Searcy and MacInnis (1970a and 1970b).

# **Biochemical Estimation of Digestive Enzyme Activity**

# Preparation of enzyme extract:

The enzyme extract preparation was carried out according to the method of Applebaum et al., (1964) with some modification. Only midgut was taken from both larvae and pupae, kept in ice-cold insect Ringer's solution, accurately weighed and homogenized for 3 minutes at 0°C in ice-cold citrate phosphate buffer (pH-6.8) using pestle and mortar. The midgut was suspended in icecold buffer and made up to 1 ml, the homogenate was centrifuged at 10,000 rpm for 15 minutes and the supernatant was used as an enzyme for estimation of digestive enzyme activity. The enzyme activity of amylase and invertase were estimated by the methods of Ishaaya and Swirsky (1970), while protease activity by Snell and Snell (1971) method and lipase activity by the method of Cherry and Crandel (1932).

# Preparation of midgut for bioassay experiment:

Alimentary canal was dissected out in insect saline, the midgut was separated,

contents of the midgut were removed by injecting the insect saline into the open midgut tube and contents were flushed out. The epithelial tissue was washed in insect saline and transferred to fresh saline. The extract of the midgut epithelia was prepared, the homogenate having a concentration equivalent to two midgut epithelia / 10 ml saline was bioassayed for its effect on in vitro digestive enzyme secretion in preparation of the midgut. The midgut preparation was incubated with 2 ml of incubation solution (midgut epithelial extract) in the bioassay apparatus for 30 minutes with a bubbling gentle stream of oxygen. After incubation, the midgut preparation was taken out and washed in insect saline, the gut was opened and contents were collected in 0.5 ml distilled water for estimation of protease and amylase activity. In control experiments, ligated midgut tubes were incubated in insect saline.

## **SDS-PAGE Electrophoresis**

Midgut extraction was carried out by the method of Laemmli, (1970) with some minor modifications (Barsagade, 1998). The molecular weight of the protein bands was estimated with the help of Gel.Doc.

## Results

## **Biochemical Analysis**

Concentration of the biomolecules present in the midgut epithelial cells of larvae and pupae were found as-

## **Total DNA concentration**

The total midgut DNA concentration in fifth instar was about 0.58  $\pm$  0.08  $\mu g/mg$ , it decreased to 0.3  $\pm$  0.06 $\mu g/mg$  in early pupa. The total DNA concentration thereafter increased gradually up to 0.45  $\pm$  0.04  $\mu g/mg$  and 0.66  $\pm$  0.08  $\mu g/mg$  in mid pupa and late pupa respectively (Fig.1).

#### Total RNA concentration

The total RNA concentration in the fifth instar larvae was estimated as  $12.2 \pm 0.99 \,\mu\text{g/mg}$  mg while it was  $7.12 \pm 0.59 \,\mu\text{g/mg}$  in early pupa, increased to  $9.12 \pm 1.01 \,\mu\text{g/mg}$  in mid pupa and was found to be  $18.6 \pm 1.01 \,\mu\text{g/mg}$  in the late pupal stage (Fig-2).

### **Total Protein concentration**

The total midgut protein concentration in fifth instar was about 4.5  $\pm$  0.29  $\mu$ g/mg. It decreased to 2.04  $\pm$  0.22 $\mu$ g/mg in early pupa, the total protein concentration thereafter increased gradually up to 3.73  $\pm$  0.31  $\mu$ g/mg and 6.5  $\pm$  0.14  $\mu$ g/mg in mid pupa and late pupa respectively (Fig-3).

## **Total Carbohydrate concentration**

The total carbohydrate concentration in the fifth instar larvae was estimated as  $0.70 \pm 0.0046 \mu g/mg$  while it was  $0.67 \pm 0.0023 \mu g/mg$  in early pupa, increased to  $0.68 \pm 0.0021 \mu g/mg$  in mid pupa and was found to be  $0.72 \pm 0.0041 \mu g/mg$  in the late pupal stage (Fig-4).

# Midgut Digestive Enzyme

The digestive enzymes, amylase, invertase, protease, and lipase in the midgut have been demonstrated qualitatively and quantitatively. Enzyme estimation in the larval and pupal stages is summarized in table 1. and fig.5.

Table-1: Enzyme estimation in larval and pupal stages of Apis cerana indica. (Mean ± SD)

S. No.	Enzyme	V-instar µg/mg	Early Pupa µg/mg	Mid Pupa μg/mg	Late Pupa µg/mg
1.	Amylase	0.19± 0.0054	0.53± 0.0187	0.028± 0.022	0.32± 0.014
2.	Invertase	0.34± 0.015	0.44± 0.0122	0.77± 0.015	0.80± 0.055
3.	Protease	0.75± 0.0057	0.78± 0.01	0.43± 0.0291	0.82± 0.021
4.	Lipase	4.5± 0.05	5.5± 0.291	2.9± 0.212	3.2± 0.254

# **Effect of Midgut Extract**

Bioassay experiment was conducted to study the effect of midgut extract on the midgut amylase and protease activity in the fifth instar larvae of honey bee *Apis cerana indica* (Fig.6). During the bioassay experiment the midgut extract showed the significant (P<0.0001) effect elevating amylase and protease activity in fifth instar.

# Amylase activity:

The midgut amylase activity was measured about 0.71 ± 0.089 mg glucose/

midgut /minute after the incubation in midgut extract for 30 minutes while, it was observed  $0.51 \pm 0.071$  mg glucose/midgut/minute in the normal condition.

## Protease activity:

The midgut protease activity was measured about 0.94 ± 0.091 mg protein/midgut/minute after incubation in midgut extract for 30 minutes while, it was noticed about 0.79 ± 0.083 mg protein/midgut/minute in normal condition.

## **Electrophoretic Analysis**

The SDS PAGE electrophoretic analysis of midgut extract demonstrated about eleven bands of protein among which six bands were predominant ranging from 26-65 kDa molecular

weight in first to fifth instar larvae, early, mid and late pupae of *Apis cerana indica*. While an additional band of 12KD protein was found only in late pupa (Fig. 7).

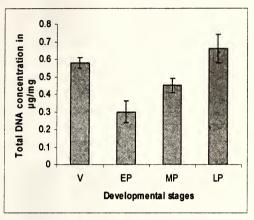


Fig. 1 Total DNA concentration during post-embryonic development

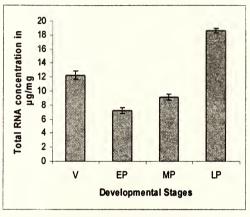


Fig. 2 Total RNA concentration during post-embryonic development.

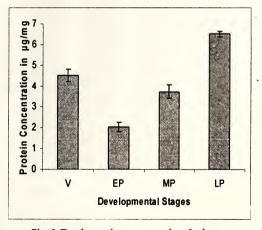


Fig. 3 Total protein concentration during post-embryonic development.

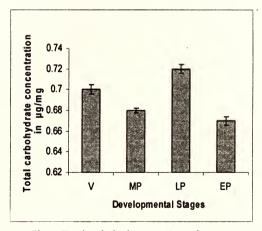


Fig. 4 Total carbohydrate concentration during post - embryonic development.

Abbreviations : V – Vth Larval stages ; EP- Early pupa ; MP – Mid pupa ; LP –Late pupa

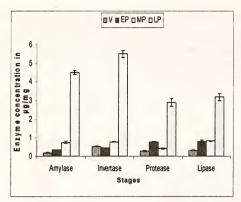


Fig. 5 Estimation of enzymes in larval and pupal stages during post-embryonic development.

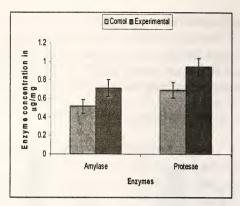


Fig. 6 Effect of midgut extract on midgut activity in V-instar larva.

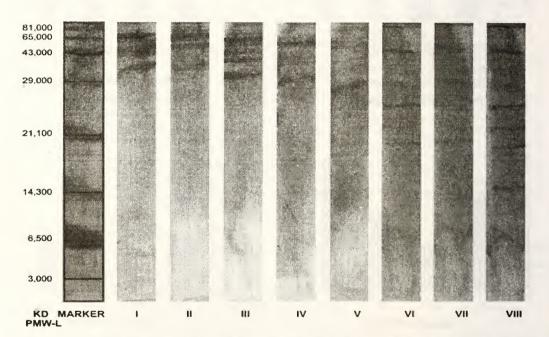


 Fig. 7 : SDS-PAGE analysis of the midgut extract of larvae of Apis cerana indica showing the presence of protein pattern.

Abbreviation: I- first instar, II- second instar, III - third instar, IV - fourth instar and V- fifth instar, VI-early pupa, VII-mid pupa, VIII-late pupa.

## Discussion

During the post embryonic development, the differentiation of midgut epithelial cells occurs in honey bee (Chapman, 1985a and 1985b and Cruz-landim and Cavalcante, 2003) while in the successive development. replacement of larval midgut epithelial cells to adult by formation of new epithelium is found in Apis cerana indica (Barsagade and Kelwadkar. 2008). In the last larval instar the midgut epithelial cells, produce various digestive enzymes that help for the food digestion in Apis mellifera (Cavalcante and Cruz-Landim, 2004). During larval development midgut cells are replaced by the other cells depending upon death of larval digestive cells, proliferation of regenerative cells occurs in order to constitute a new digestive epithelium in the adult, Apis mellifera (Geogorc and Bowen, 1997) and in Apis cerana indica, (Barsagade and Kelwadkar, 2008).

The midgut columnar epithelial cells containing the biomolecules such as DNA, RNA and Proteins are actively engaged in the protein synthesis in order to secure various digestive enzymes during the larval-pupal metamorphosis (Fuji, 1979, Wigglesworth, 1972 and Chapman, 1998).

In Apis cerana indica, total DNA, total RNA, total proteins and total carbohydrates were intensely demonstrated in the nuclei and perikarya of columnar epithelial cells, as found in Apis florea and Apis mellifera, described by earlier workers showing their role in protein synthesis (Pearse, 1968; Ban and Prezlec, 1974). The similar role of these biomolecules may be played in the protein synthesis during metamorphosis in Apis cerana indica.

Various biochemical tests reveal the presence of amylase, invertase, protease and lipase activity in the midgut of larvae of *Apis cerana indica* which are very specific in their

activity (Horie, 1970, Banergee and Saxena, 1983, Wajiro et al., 1984 and Zufelato et al., 2004). The higher level of enzyme acid phasphatase activity in the larval peritrophic membrane in Apis mellifera is mostly used for digestion (Cavalcante and Cruz-landim, 2004). Similarly, activity of other fourteen enzymes was determined in the midgut of Apis mellifera and these enzymes were alfa-amylase, 4 alfaglucosidase, 3 protease, 5 amino peptidase, lipase etc. (Banerjee and Sexena, 1983). According to Mahmoud (2007), the high food metabolism level depends on the protein concentration in the midgut and haemolymph. it may reflect the digestion level with acid phasphatase and other digestive enzymes. The present investigation reveals that the concentration of biomolecules viz. total DNA. total RNA, total protein and total carbohydrates in the fifth instar larvae of Apis cerana indica. shows initial depletion and then a significant rise because of involvement of total protein consumption in the midgut for synthesis of digestive enzymes.

The study also supports the findings of earlier works, showing the amylase, invertase, proteases and lipase activity during the larval-pupal metamorphosis in *Apis cerana indica*. Cavalcante and Cruz-landim (2004), noticed the mass range of different proteins varying form 19 to 142 kDa, increased greatly from larvae to pupae and tends to decrease during pupation, until the phase of brown eye pupae and increased again in the pupae with pigmented body (pharate adult). In *Apis cerana indica*, 11 bands were observed, among which six bands were predominant ranging from 26–65 kDa, found in the midgut extract of developing larvae and pupae, supporting the earlier studies.

## References

- Applebaum, S. W., Harper, I. and Bondi, A. 1964. Amylase secretion in the larvae of Prodenica litura F. Insecta. Comparative biochemistry and Physiology 13: 107-111.
- Ban, A. and Przelec 1974. The influence of starvation on ribonucleic acid and protein synthesis in midgut epithelium of Gallaria mellonella larva. Folia Histochemica et Cytochemica 11(314): 177-183.
- Banergee, A. and Saxena, P. 1983. Digestive enzymes in the alimentary canal and associated glands of foragers of Apis cerana indica. Lucknow: Department of Zoology, University of Lucknow.
- Barsagade, D.D. 1998. Studies on the silkgland development and silk protein syntheisis in tasar silkworm, Antheraea mylitta(D). Ph.D. thesis, Nagpur University, Nagpur, India.
- Barsagade, D.D. and Kelwadkar, M. K. 2008. Ultrastructural changes in the midgut during metamorphosis in Apis cerana indica. Journal of Industrial Entomology 6(1): 29-35.
- Cavalcante, V.M. and Cruz-landim, C. 2004. Electrophoretic protein pattern and acid phosphates activity in the midgut extract of Apis mellifera L. (Hymenoptera: Apidae) during metamorphosis. Neotropical Entomology 33 (2):169 -172.
- Chapman, R.E. 1985a. Co-ordination of digestion. In:
  Kerkut, G.A. and Gilbert, L.I. (eds.).
  Comprehensive Insect Physiology,
  Biochemistry and Pharmocology. Pregaman
  Press, Oxford 4: 213-240.
- Chapman, R.E. 1985b. Co-ordination of digestion. In:
  Kerkut, G.A. and Gilbert, L.I. (eds.).
  Comprehensive Insect Physiology,
  Biochemistry and Pharmocology. Oxford,
  Pregaman Press, 4:165-211.
- Chapman, R.E. 1998. Endocrine system, A text book of insect structure and function IV. Cambridge: Cambridge University Press.
- Cherry, I. S. and Crandell, L. A. 1932. Determination of

- serum lipase activity. Annual Journal of Physiology 100: 266.
- Cruz-landim, C. and Cavalcante, V. M. 2002. Cell nucleus activity during post-embryonic development of Apis mellifera L. (Hymenoptera: Apidae). Genetic and Molecular research 1(2): 131-138.
- Cruz-landim, C. and Cavalcante, V. M. 2003. Ultrastructural and Cytochemical aspects of metamorphosis in the midgut of Apis mellifera L. (Hymenoptera: Apidae: Apinae). Zoological Science 20(9): 1099-1107.
- Dubois, M., Giller, K. A., Hamilton, J. K., Rebers, P.A. and Smith, F. 1956. Colorometric method for determination of sugar and related substances. Analytical Chemistry 28:350.
- Fuji, H. 1979. Studies on function and structural conversion of the midgut during larval, pupal, transition in silkworm, Bombyx mori (4). Hormonal regulation of metamorphosis. Science Bulletin of Faculty of Agriculture Kuyushu, University 33 (2-3): 127-166.
- Gregorc, A. and Bowen, I.D. 1996. Scanning electron microscopy of the honey bee (Apis mellifera L.) larvae midgut. Zb Vet. fAk.-Univ.Ljubl. 33: 261-269.
- Gregorc, A. and Bowen, I. D. 1997. Programmed cell death in the honey-bee (Apis mellifera L.) larvae midgut. Cell Biology International 21(3): 151-158.
- Horie, K. 1970. Physiological studies on the digestive tract of larvae of the Silkworm, Bombyx mori L. Il Carbohydrates in the digestive juice and midgut. Bulletin of Comparative Sericulture 15: 365-380.
- House, H. L. 1974. Physiology of Insecta. In: Rockstein, M. (ed.). International Conference on Apiculture in Tropical Climates, Division Entomology IARI, New Delhi: 367-371.
- Ilshaaya, I. and Swirsky, E. 1970. Invertase and amylase activity in armored scales Chrysomphalus aoridam and Aonidiella auranti. Journal of Insect Physiology 16: 1599-1606.

- Laemmli, U.K. 1970. Cleavage of structural protein during the assembly of head of bacteriophage T<sub>4</sub>. Nature 227: 680.85.
- Lowry, L. H., Rosebrough, N.J., Farr A.L., and Randall, R. J. 1951. Protein measurement with folin-phenol reagent. Journal of Biological Chemistry 2193: 65-75.
- Mahmoud, E. Z. 2007. Factor affecting on the food metabolism in some honey bee races. Journal of Applied sciences Research 3(4): 311-316.
- Neves, C. A., Gitirana, L. B. and Serrao, J. E. 2003. Ultra Structure, of the midgut endocrine cells in Melipona quadrifascita anthidioides (Hymenoptera: Apidae). Brazilian Journal of Biology 63: (4).
- Pearse, A.G. 1968. Histochemistry-Theoritical and Applied. London: J. and A. Churchill Ltd.
- Prabhu, V. K. K. and Sreekumar, S. 1994. Endocrine regulation of feeding and digestion in insects. In: Agrawal, P. (ed.). Entomological Research, Scientific Publishers, Jodhpur: 117-135.
- Roth, T. F. and Porter, K. R. 1964. Yolk protein uptake in oocyte of mosquito Aedes aegypti L. Journal of Cell Biology 20: 312-332.
- Searcy, D.G. and MacInnis, A. J. 1970a. Biochemical estimation of DNA by Burton's Diphenyamine method. Biochemical Journal 62: 215-223.
- Searcy, D.G. and MacInnis, A. J. 1970b. Biochemical estimation of RNA by Dische-Orcinol method.

- Biochemical Journal 62: 315-323.
- Snell, F.D. and Snell, C.T. 1971. Colorimetric method of analysis. New York: 3<sup>rd</sup> Edition Academic Press
- Snodgrass, R.E. 1935. Principle of insect morphology. New York: New York Press.
- Snodgrass, R. E. 1956. Anatomy of honeybee. New York: Comell University Press.
- Sunitha, V.B., Reena, T., Harshini, S. and Sreekumar, S. 1999. Is gut pH regulated by midgut endocrine system. Indian Journal of Experimental Biology 37: 423-426.
- Wajiro, M., Fuji, H. and Sakaguchi, B. 1984. Activity and polymorphism of digestive juice amylase in various strains of Bombyx Mori. Journal of Sericulture Sciences of Japan 55(6): 496-500.
- Wigglesworth, V. B. 1972. The principles of insect physiology 7th Ed. New York: N. Y. John Wiley and Sons.
- Wigglesworth, V. B. 1977. The principles of insect physiology, London: Methuen Press.
- Zufelato, M. and Laureno, A. P., Simoes Z. L., Jorge, J.A., Bitondi, M. M. 2004. Phenoloxidase activity in Apis mellifera honey bee pupae, and ecdysteroid dependent expression of the prophenoloxidase m-RNA. Insect Biochemistry and Molecular Biology 34(12): 125-68.